Review of parasitic zoonotic infections in Egypt Mousa A.M. Ismail, Ahmed H.A. Eassa, Abeer M.A.I. Mahgoub, Nadia El-Dib

Department of Medical Parasitology, Faculty of Medicine, Cairo University, Cairo, Egypt

Correspondence to Ahmed H.A. Eassa, MBBCH, MD, Faculty of Medicine, Cairo University, Cairo, Egypt. e-mail: ahmegy@hotmail.com

Received 21 November 2018 Accepted 13 December 2018

Kasr Al Ainy Medical Journal 2018, 24:91–100

Zoonoses are diseases and infections that are transmitted in nature between vertebrates and humans. Zoonoses consist of an interaction among at least three species: one pathogen and two hosts - animals and humans. This review aimed at shedding in depth light on zoonotic parasitic diseases in Egypt, with special reference to their relative incidence between humans, reservoir animals, sources of human infection and control policies. According to the available literature, many parasitic zoonoses are endemic in Egypt. In rural areas, intestinal parasitic zoonoses are widespread and are the leading cause of diarrhea, particularly among children. Some parasitic zoonoses are mainly found in certain areas in Egypt, for example, cutaneous leishmaniasis and zoonotic babesiosis in Sinai. Other locations in Egypt have a history of certain parasitic zoonoses, such as visceral leishmaniasis in the El Agamy area in Alexandria. Fortunately, control programs have led to a dramatic decrease in the prevalence of other zoonoses, such as intestinal schistosomiasis and fascioliasis in the country. In Egypt, animal reservoirs of parasitic zoonoses have been identified. These include rodents, stray dogs, and cats as well as domestic and farm animals and birds. Many vectors have also been revealed, typically mosquitoes and ticks, which pose real threats for disease transmission. Strict control strategies are needed to upgrade and complement current efforts at eradicating parasitic zoonoses in Egypt.

Keywords:

arthropods, Egypt, helminth, protozoa, zoonoses

Kasr Al Ainy Med J 24:91–100 © 2019 Kasr Al Ainy Medical Journal 1687-4625

Introduction

Zoonotic diseases are diseases that may be transmitted between animals and humans. It is a Latin name meaning ZOON=Animal and Noson=Disease. An estimated 60-70% of infectious diseases in humans globally are zoonotic, and great percentages originate from wildlife [1]. Approximately 25% of the world's population may be suffering from parasitic diseases. These infections are classically predominant in underdeveloped agricultural and rural areas of tropical and subtropical regions, causing reduced worker efficiency and a waste of economic resources. As many as 400 million people of the Middle East and the North Africa region including Egypt may be affected due to environmental, social, educational, and economic factors [2,3]. In general, the consequences of such zoonoses are amplified because they lead to morbidity in both immunocompromised and immunocompetent patients [4].

Intestinal parasites are common in school-age children because of overcrowding and behavioral patterns that contribute to the extent of parasitic infection. High incidence rates of parasitic infection among children have been documented, with levels reaching up to 48% [5,6]. The medical and financial impacts of parasitic zoonoses have been studied by many scientists in Egypt. The hazards of zoonoses are significant due to the proximal vicinity of cattle and other domestic animals to households, specifically in rural parts, and, similarly, pets and stray animals, especially canines and felines, are predominant all over the country [3,7]. Numerous surveys of parasitic infections accomplished in rural areas of Egypt conveyed high infection rates of solitary and numerous gastrointestinal parasitic infections, with levels reaching up to 85% [8,9]. Indeed, both domestic and wild animals and birds are well-documented reservoir hosts in Egypt. This is predominately noted because of the vast unpopulated desert areas where wild rats, felines and canines live and replicate. An epidemiological study of wild rodents described an infection rate of parasites of 54% (93/ 172), including 28% cestodes, 7% nematodes, 8% Acanthocephala, and 41% protozoa [3]. In addition, a cross-sectional study revealed that the overall incidence of helminths in domestic rodents in Dakahlia was 53%,

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

and a total of 24 species of helminths were recognized [3,10,11].

Arthropod-borne zoonotic parasites are found in Egypt; the role of arthropods as vectors for some parasitic infections is well documented, like the sandfly vector of leishmaniasis, or like ticks, the vectors of Babesia spp. infection. In addition, nonblood-sucking arthropods play a significant role in the mechanical spread of human parasites such as the American cockroach (Periplaneta americana) and the house fly Musca domestica var. vicina [12]. Nevertheless, apparently due to lack of vectors and other environmental elements, numerous severe parasitic diseases such as Kala-azar and sleeping sickness are not endemic in Egypt, in contrast to tropical Africa. However, several zoonotic parasitic infections are present in Egypt, such as soiltransmitted nematode infections, hydatidosis, filarial infections, schistosomiasis, fascioliasis and leishmaniasis [3]. For example, a cross-sectional study was planned to investigate the incidence, control policies and public health importance of the gastrointestinal parasites of sheep from the Nile Delta, Egypt, and showed the existence of parasites in more than 60% of the cases [13].

According to the WHO, an obvious modernization in drinking water sources and sanitation amenities has occurred in Egypt, with over 90% of the population having access to healthier drinking water sources. Furthermore, the wellbeing of people has improved significantly compared with the regional average; consequently, along with prevention and control efforts, infections such as schistosomiasis and fascioliasis have decreased among humans and animals in the country [3].

Zoonotic parasites can be categorized into four classes: direct zoonotic, metazoonotic, cyclozoonotic, and saprozoonotic. Direct zoonotic parasites infect humans directly from animals and include Entamoeba histolytica, Cryptosporidium parvum, Toxoplasma gondii, and Sarcoptes scabiei. Metazoonotic parasites comprise Fasciola spp. and Schistosoma spp., and these require a molluscan/invertebrate intermediate host to infect humans. In contrast, cyclozoonotic parasites have vertebrate intermediate hosts and include Echinococcus granulosus, Taenia saginata, and Taenia solium. However, saprozoonotic parasites can infect humans from soil or water, such as Ancylostoma caninum and Strongyloides stercoralis [3,14]. Bearing in mind the dynamic alterations in the Egyptian health, ecological, and veterinary sections, which readily affect reservoirs of parasitic infections and linked zoonoses, this review aimed to cover such infections and deliver an updated insight on zoonotic parasites in the country.

Zoonotic protozoan infections in Egypt Intestinal zoonotic protozoal infections

Gastrointestinal protozoa are one of the main health problems with a high predominance all over the world. This prevalence depends on several factors including environmental, social, and economic ones. All in all, a high incidence of intestinal parasites is often correlated with several health problems in a given society. This is predominately seen in low-income nations where gastrointestinal parasites in children are linked to inhibition of normal growth, low intellectual progression, and vitamin deficiency by malabsorption, chronic diarrhea, and dysentery [15,16]. Definitely, in Egypt, parasites are the major cause of diarrhea, with the prevalence reaching over 60% [17–19].

The majority of parasitic gastrointestinal problems in Egypt was caused by protozoa (57.6%); whereas helminthes were found only in 9.9% of cases [20]. Further studies in the Delta region revealed that up to 67.1% of chronic diarrheic patients had parasitic infections; involving mixed infections in 12.9% of total chronic diarrhea patients. It is noteworthy that both immunosuppressed and immunocompetent patients showed a high prevalence of intestinal protozoa disease in Egypt [4,21].

In Egypt, *E. histolytica* was reported to have a prevalence that ranged from 0 to 57% in diarrheic patients, with greater infection rates in poor regions and Upper Egypt [4,21,22]. As for probable reservoir animals, amebiasis has been described in both wild and domestic animals [11].

Giardiasis produced by *Giardia intestinalis* is a major international diarrheal disease. It is a predominately common intestinal parasite in Egypt, particularly among children. *G. intestinalis* infection rates of 10–34.6% have been stated. Prevalence rates of 11 and 15.4% among children were reported, whereas, among chronic diarrhea patients, the prevalence reached up to 27.3% [3,6,23]. In contrast to former reports abroad declaring that assemblage B is the most frequent genotype of *G. intestinalis* (80%) [24], it was later reported that the greatest prevalent genotype in Egypt was assemblage A [25]. Other assemblages, C and E, have also been noted [26]. Concerning reservoir animals in Egypt for *Giardia* spp., cysts were revealed in stool samples of 2% of homeless cats [27] and 8% of wild rats [10]. Furthermore, fish have been incriminated as a potential reservoir for G. *intestinalis*, as the parasite has been found in fish, with a prevalence of 3.3% [28].

In Egypt, infection with Cryptosporidium spp. is one of the growing reasons for diarrhea, with infection rates up to 49% between both inpatient and outpatient clinic attendants [3,18,29]. More specifically, it was demonstrated that infection with C. parvum protozoan was the most common in immunosuppressed patients, with an infection rate of 60.2% [25]. Cryptosporidiosis, including C. hominis and C. hovis, is a well-documented zoonotic disease, and has been described in farmers and their domestic animals [3], with a prevalence rate of 32.2% in ruminants [29]. Calves drinking from canals or underground water were at a higher risk of infection than calves drinking tap water [29]. Oocysts have been identified in the fecal samples of neonatal calves (30.2%) and neonatal lambs (30%). Different genotypes have been identified in animals with a high incidence of zoonotic C. parvum subtype families (IIa and IId) in Egypt [29]. In animals, C. parvum appears to be the dominant species with infection rates of up to 82.8% [29,30] in Egypt. There have been reports recording the existence of additional Cryptosporidium spp. in Egyptian domestic animals: C. ryanae (11.8%), C. bovis (4.1%), and C. andersoni (6.9%) [30]. In addition, Cryptosporidium spp. was detected in wild rats, with a prevalence of 22.7% (C. paroum) and 20.3% (C. muris) [3].

Sarcocystis spp. is a zoonotic parasite of livestock animals. Moreover, S. cruzi, S. hirsute, and S. hominis have canids, felids, and humans as definitive hosts, correspondingly, and can affect bovines as intermediate hosts creating muscle cysts [3,31]. This parasite is endemic in Egypt; cysts were repeatedly identified in cattle and buffalo during meat inspection. Furthermore, Sarcocystis spp. infection in stray cats was reported at a rate of 1% [32]. Thus, stray cats may, consequently, be potential reservoirs in Egypt, too.

Blastocystis hominis has been revealed in Egypt with a prevalence rate of 22.4% in asymptomatic patients, compared with 12.1% in patients with diarrhea and in immunosuppressed children [4]. Its zoonotic potential is not largely explored, but an infection rate of 3% was reported among stray dogs in Egypt [7]. This suggests a possibility that *B. hominis* may be a zoonotic infection in the country.

In Egypt, numerous additional zoonotic intestinal protozoa have been reported [33] with variable

infection rates, such as Iodamoeba butschlii (16%), Isospora hominis (7.7%), Endolimax nana (6.9%), Entamoeba hartmani (5.9%), Dientamoeba fragilis (5.1%), Chilomastix mesnili (5.1%), Trichomonas hominis (4.2%), Microsporidia spores (3.2%), Enteromonas hominis (1.9%), and Embadomonas intestinalis (1.3%).

Blood and tissue zoonotic protozoal infections

Leishmaniasis is a vector-transmitted parasitic infection with a worldwide distribution. Infection with Leishmania spp. has a wide range of clinical manifestations, ranging from skin infections to lethal visceral disease. The Sinai Peninsula in the northeast part of Egypt, a sporadically inhabited locality, has seen occasional cases of zoonotic cutaneous leishmaniasis. The vectors of Leishmania spp. such as Phlepotomus papatasii and P. sergenti sandflies have been identified in endemic regions [34]. Previous studies on the etiologic factor of cutaneous leishmaniasis have steadily recognized Leishmania major and L. tropica as the primary and secondary causes, correspondingly. Wildcaught rodents were identified as reservoir hosts of Leishmania spp. infection in endemic areas. Gerbillus pyramidum floweri have been documented to be naturally infected with L. major and L. tropica in North Sinai [35]. Additional rodent species, Rattus norvegicus, was classified as a reservoir host of Leishmania infection in nonendemic regions of Qualyobia [36].

L. donovani producing visceral leishmaniasis has been reported in Egypt, mostly at the Mediterranean coast in Alexandria Governorate; where 27 cases were discovered from 1982 to 1985 through active and passive case detection. Of them, 22 cases were revealed in El Agamy, a resort town of 50 000 inhabitants situated 15 km west of the city of Alexandria. An additional case of infantile visceral leishmaniasis was recognized in an adult farmer in Banha city. Likewise, infrequent cases were reported among hypersplenic patients in Dakahlia Governorate. No new cases have been discovered since these sporadic cases. In contrast, the preceding history of visceral leishmaniasis indicates an ongoing possibility for such infections in Egypt. Dogs are a chief reservoir host for visceral leishmaniasis, including in El Agamy, Alexandria, where L. infantum was detected in stray dogs [37]. An epidemiological study in Giza confirmed that antibody to Leishmania spp. was detected in five of 50 (10%) dogs examined for Leishmania causing visceral leishmaniasis [38].

Toxoplasmosis trigged by *T. gondii* infection is a rising global health concern, with up to one-third of the world's population assessed to be infected with the

parasite [39]. Many authors revealed high rates of seroprevalence of *T. gondii* in Egypt. For example, among 260 asymptomatic blood donors, 155 (59.6%) were anti-*Toxoplasma* immunoglobulin (Ig)G positive, as detected by enzyme-linked immunesorbent assay (ELISA) [40].

Toxoplasmosis causes a diversity of effects, particularly in pregnant women facing a possible risk of fetomaternal transmission leading to abortion, premature birth, and other congenital deformities of the newborn. The hazard of fetomaternal transmission of Toxoplasma spp. has been documented with antibody prevalence of 51.5% of pregnant women, as revealed by ELISA. Seropositivity to specific anti-Toxoplasma IgG antibodies was 57.9, 58.1, and 44.7% in serum samples from haphazardly collected samples, full-term pregnant women, and women who had suffered a respectively [41]. miscarriage, However, the seropositivity rates to specific anti- Toxoplasma IgM for the same groups were 10.5, 6.5, and 23.7%, respectively. Moreover, another study had previously described that 10 of 42 meningoencephalitis patients (26%) had Toxoplasma IgG antibodies in cerebrospinal fluid samples, evaluated by immunofluorescent antibody assay [42]. High infection rates of T. gondii infection among cats and other animals have been reported. The incidence of T. gondii varied between 9% and up to 95.5% among Egyptian cats by modified agglutination test, which implies a high risk of T. gondii human disease [43]. In domestic animals, high levels of antibodies have been demonstrated. Immunofluorescent antibody was revealed in the serum of slaughtered animals at a prevalence of 48.8%. Anti-Toxoplasma antibodies were reported using a modified agglutination test in 59.5% of domestic turkeys, 47.2% of chickens, and 50% of ducks, in addition to 10.8% of cattle, as demonstrated by ELISA [44].

Concerning other uncommon blood and tissue zoonoses in Egypt, zoonotic *Babesia* infections were reported in Sinai, where rodents are *Babesia microti* reservoirs, particularly the mice of *Acomys* spp., whereas *Ixodes ricinus* ticks have been detected to be the vectors [45,46]. *Neospora caninum* is a predominant animal protozoan parasite with a global distribution. It is not identified to produce human infection; however, antibodies to *N. caninum* were recognized in 7.9% of pregnant women in a study in Egypt, whereas the detection rate was 20.4% in cattle and 1.9% in rabbits [41]. In another rare incidence, a case of zoonotic infection of *Trypanosoma evansi* was described [47].

Zoonotic trematodal infections in Egypt

In Egypt, animal and human fascioliasis is an endemic disease produced by Fasciola spp. It has almost reported in all governorates, been particularly the Nile Delta and Alexandria [48-50]. Fascioliasis is considered hyperendemic in the Delta region [51], with a frequency of 4.8% of outpatients examined in that study. However, fascioliasis spreads even as far west as the desert oases [52]. Over the last three eras, cases of human fascioliasis have been reported. For example, a prevalence of 5.2–19.0% (mean=12.8%) was documented in the Nile Delta villages [51]. Afterwards, human infections have begun a fading drift in Egypt [48–50,53]. However, more recently, it was stated that it may be re-emerging in upper Egypt [54].

Donkeys' and camels' infection with *Fasciola gigantica* has been recorded in Egypt, in addition to that of grassgrazing domestic and farm animals. Fascioliasis was recognized to be endemic in sheep [55], particularly in the Nile Delta, as identified by abattoir surveys (20.6%) and by the microscopic revealing of *Fasciola* spp. in fecal samples (12.7%) [56].

Heterophyiasis is an intestinal fluke infection that is endemic in Egypt, particularly in brackish and fresh water fish in northern areas, where the parasites are indigenous in the lakes and neighboring regions. The peak rate of infection was found in fishermen (33.8%) and native inhabitants in northern Egypt (13.3%) [57,58]. The overall prevalence of heterophyid infection of fish was documented to be 32% (the mean of 22% in brackish water fish and 42% in fresh water fish) [58]. Moreover, the prevalence of heterophyid metacercaria in fresh water fish was recognized to be 95.4% in Ismailia [41]. Several heterophyids have been recognized from fish in Egypt. Adult heterophyids from *Heterophyes* heterophyes, Heterophyes aequalis, Pygidiopsis genata, Haplorchis yokogawai, Prohemostomum vivax, Phagicola ascolonga, and Stictodora tridactyla were recorded from encysted metacercaria-fed puppies. Moreover, the prevalence of Heterophyes heterophyes was 3% among stray cats in Kafr Elsheikh in the northern area of the Delta [27,59].

Schistosomiasis is the third leading endemic parasitic disease in the world. It was stated that more than 200 million persons in 74 countries are infected and that 120 million have symptoms of the disease. In 2011, 42 countries in Africa, including Egypt, were considered Schistosoma hematobium rates declined from 60 to 70% in 1925 to 5% in 1996, and *S. mansoni* rates decreased from 32% in 1932 to 12% in 1996 [60]. In recent years, control programs are applied, and the prevalence of infection with *Schistosoma* spp. has declined, particularly with the continuous and reliable praziquantel treatment.

Subsequently, lower infection rates ranging between about 1 and 5% have been recorded for *S. mansoni* in Egypt [9,11,20]. In Egypt, the incidence and species dissemination of schistosomiasis vary classically in Upper verses Lower Egypt. A study executed in nine governorates reported that *S. mansoni* was uncommon in Upper Egypt, excluding Fayoum, with an incidence rate of 4.3%. Yet, the incidence of *S. mansoni* in five governorates in Lower Egypt, where it is endemic, had an average of 36.4%. The prevalence of *S. haematobium* in four governorates in Upper Egypt, where it is also endemic, was revealed to have an average of 7.8%. This species is rare in the Delta areas [62].

Conflicts and human and animal migrations are key social factors in prevention, control, or eradication of zoonotic parasitic infections in Egypt, while local political motivation, reinforced global and intersectoral cooperative efforts for surveillance, mass drug administration, and vaccination are vital for eradication [63,64]. However, a novel report revealed rather active S. mansoni transmission status in certain districts in Lower Egypt, using the urinecirculating cathodic antigen and Kato technique [65]. In this report, of 35 regions surveyed in five governorates, S. mansoni infection was documented in 19 (54.3%) districts using Kato technique, and in 31 (88.6%) districts by the urine test. Male individuals and higher age groups had significantly higher urine antigen prevalence rates. On the basis of these discoveries, authorities of the Ministry of Health and Population implemented a new eradication strategy by readapting thresholds for mass treatment with praziquantel and targeting all transmission regions [3,65,66].

It is noteworthy that the zoonotic potential of schistosomiasis in Egypt was emphasized before, as *S. mansoni* and *S. haematobium* were detected as a natural double infection in the Nile rat, *Arvicanthis niloticus*, from a human endemic region in Egypt [67].

Zoonotic cestodal infections in Egypt Intestinal zoonotic cestode infections

Taeniasis due to infection with T. saginata is more in Egypt than due to infection with T. solium, as the intermediate host of the latter is pig, and pork in not broadly consumed for religious reasons. However, few studies are reported about taenisais in Egypt. For example, an infection rate of human T. saginata infection of 1.1% was reported [11]. Another study was carried out at an abattoir, on meat samples from 6 434 039 slaughtered animals, between 1994 and 1997; the infection rate was 0.2% in domestic cattle, 7.3% in imported cattle, 0.1% in buffaloes and 0.1% in pigs, with an overall incidence of cysticerci (Cysticercus bovis and cysticercus cellulosae) of 0.7% [68]. Although human cysticercosis due to consumption of *T. solium* egg is not a health problem in Egypt, it has a harmful effect on intermediate host animals and remains a potential zoonosis source. It was demonstrated that 20% of the studied cattle were infected with cysticercus bovis, and 12% of pigs were infected by cysticercus cellulosae, among slaughtered animals in El-Minia in Upper Egypt [69].

Hymenolepis nana infection remains a common intestinal cestode disease, particularly among children in Egypt, whereas H. diminuta is an unusual zoonotic parasite. Earlier studies of H. nana infections in children specified a high prevalence of 16% [70]. In another report on outpatients of a hospital in Dakahlia, the reported rate of H. nana in stool samples was 3.9% [11]. In Menoufia, among 2292 farmers, the rate of reported H. nana eggs in stool samples was 3% [9]. It was demonstrated that *H. nana* infected up to 16% of children registered in their report at Fayoum University Hospital [71]. As for reports on H. diminuta infections, a lower rate of 1.4% was detected [11]. Rodents are considered the chief reservoirs of H. diminuta infections, with an infection rate of 23.8% in Egypt [10]. More recently, an infection rate of 1.5% was found among dog populations in Egypt [7].

Concerning *Dipylidium caninum*, it was reported that it had not been described as a zoonotic infection among humans in Egypt [33]. Yet, it was reported to be at 5% in stray cats [27].

Larval zoonotic cestode infections

Cystic echinococcosis or hydatidosis due to consumption of the egg of *E. granulosus* is predominant in the Middle East and Arabic North Africa, and is endemic in Egypt [3,72]. In a

retrospective study [72] on human cystic echinococcosis in Egypt between 1997 and 1999, using 492 353 patient records, 133 (0.03%) new human cystic echinococcosis cases were reported. study seropositivity Another on levels of echinococcosis stated a rate of 5% in patients with acute and chronic hepatic diseases in Assiut and Aswan [73].

As for the incidence of *E. granulosus* in the chief final hosts in Egypt, a rate of 5% was documented among street dogs, particularly in rural areas [33]. In intermediate hosts, the overall five-year hydatidosis incidence from August 2000 to August 2005 was 2.5, 0.3, and 0.7% in camels, sheep and goats, and pigs, correspondently. However, a higher level of hydatidosis mainly in camels was confirmed, reaching up to 7.7% [74]. Six genotypes of the *E. granulosus* complex are found in Africa. In Egypt, the sheep strain (G1) was reported in sheep, goats, cattle, camels, sheep, cattle, and humans. All in all, the G6 genotype had a higher prevalence in humans and animals in the country [75].

Recently, the role of Norway rat as a potential *E.* granulosus reservoir in Cairo, Egypt, was studied [76]. Both rats and humans living in related regions were covered in that study. The results demonstrated the following overall seroprevalence rates of cystic hydatidosis in examined rats and persons: 36 and 11.9%, respectively. Cysts were recognized as *E.* granulosus hydatid cysts (G6 strain). According to the authors, this emphasizes the possible role of Norway rat in the epidemiological cycle of *E.* granulosus, particularly in urban and suburban locations.

Zoonotic nematodal infections in Egypt Zoonotic intestinal nematode infections

Capillaria spp. has been an evolving imported zoonotic infection in Egypt [77], especially in Upper Egypt governorates for the past few decades. Dissimilarly, a low prevalence of *Capillaria* spp. infection was described in the Nile Delta (1%) [20,78]. Fisheating birds are the essential host for this parasite, and man is infected accidentally [79]. Stray cats are assumed to be likely reservoir hosts, as *Capillaria* spp. was identified in them with a prevalence of 3% [27].

Previously, infection rates with *Ascaris lumbricoides* in Egypt were reasonably high, but, more recently, reports detected lower rates: under 2% [11,20]. In Egypt, dogs

are thought to be reservoir hosts of *A. lumbricoides* as environmental polluters, mounting the risk of infection in humans [80].

Anisakis simplex is not native in Egypt, and, in one study directed to fish trapped in the Red Sea, it was reported with a rate of 2.2% from the orange-spotted trevally, *Carangoides bayad* (Carangidae) fish [81].

Eosinophilic enteritis caused by *A. caninum* was once recognized in Egypt. The prevalence of IgG antibodies to this nematode in patients with vague acute or recurrent abdominal pain was found to be 11.6% [82].

Other species of nematodes have been described in dogs that may be potential reservoirs, such as *Trichostrongylus* spp. (2.6%), *S. stercoralis* (1.5%), *Enterobius vermicularis* (1.1%), and *Trichuris trichiura* (0.7%) [80]. As for *Trichostrongylus*, it is a well-known parasite of farm animals, and it was the most prevalent intestinal nematode detected in sheep in Dakahlia [83].

Extraintestinal and larval zoonotic nematode infections

In Egypt, a high prevalence of human toxocariasis denoted by anti-*Toxocara* antibodies has been verified by several investigators [84,85].

Seroprevalence of IgG antibodies was 7.7% in the general community [85], 6.2% among suspected children, and 18% in adults [84]. The reservoirs of *T. canis* infection are dogs, denoting a direct infection risk to humans [86]. A report in Egypt recorded that 56% of dogs were infected with *T. canis*, and 8% were infected with *T. leonina*. In addition, *T. cati* and *T. leonina* have been found in stray cats at a prevalence of 9 and 5%, correspondently [80]. More recently *T. canis* was detected at a much lower rate of about 5% in different dog populations in Egypt: stray, military, and domiciled [7].

Human trichinellosis due to *Trichinella spiralis* is not predominant in Islamic nations like Egypt due to religious beliefs and food behaviors. Few studies of *T. spiralis* infection in fresh and handled pork in Egypt are accessible [87,88]. As for its zoonotic potential, *T. spiralis* infection was reported in 13.3% of rodents gathered from and around slaughterhouses in Alexandria [89].

It was demonstrated that about 50 million people in Egypt and sub-Saharan Africa had bancroftian filariasis due to infection with *Wuchereria Bancrofti*. In a longitudinal report of bancroftian filariasis in the Nile Delta of Egypt, microfilaremia and filarial

antigenemia rates among 1,853 subjects more than nine years of age was 7.7 and 11.2%, correspondently, while the one-year frequency was 1.8 and 3.1%, respectively [90]. Subsequently, by examining night blood films, a high incidence of 38% was documented among asymptomatic patients with *W. bancrofti* [91]. Concerning the common Egyptian vectors, *W. bancrofti* DNA was found in 31.9% of *Culex pipiens* mosquitos gathered from an Egyptian village with low filariasis prevalence [92]. Furthermore, high rates of microfilaria were reported in mosquito vectors collected from households with significant risk factors for microfilaria spread [90,92]. In animals, *W. bancrofti* microfilaria was reported in a stray cat from Assiut in Upper Egypt [93].

Human dirofilariosis, caused by *Dirofilaria repens*, is an evolving zoonosis, and dogs are the main hosts. Three cases of human infection with *D. repens*, one pulmonary and two subcutaneous, were detected in Assiut, Upper Egypt. This was the first report of human pulmonary dirofilariosis in Africa [94]. Concerning the role of cats in the transmission of *D. immitis*, a seroprevalence rate of 3.4% was discovered in stray cats in Cairo [43]. Furthermore, a case of subcutaneous nodule in the upper eyelid was reported in Egypt due to *D. conjunctivae* [95].

Zoonotic arthropod infestations in Egypt

The worldwide mite *S. scabiei* (*Acari: Sarcoptidae*) is a compulsory ectoparasite that infests the skin of a broad range of mammalian hosts, causing sarcoptic mange in companion animals, livestock, and wildlife, as well as scabies in humans [96].

In Egypt, scabies has been detected in humans [97], wild games [98] and farm animals [99], and zoonotic bird and rat mites (Ornithoyssus) have been recognized to produce dermatitis in poultry farm laborers [100]. Rat ectoparasites that may infest humans have also been reported, comprising Xenopsylla cheopis, Hyalomma dromedarii (nymph), Echinolaelaps echidninus, and Hemolaelaps glassgowi [101]. In addition to zoonotic mites, stray dogs and cats are documented to be reservoirs for uncommon zoonotic arthropods such as Linguatula serrate [27].

Prevention and control strategies

There is a lack of information on several parasitic zoonoses control in Egypt, and, overall, there is a crucial need to magnify these surveillance strengths for most of the parasitic zoonoses. Tools exist to

control, or in some cases, eradicate such main parasitic zoonoses in Egypt. For the soil-transmitted nematode infections, coverage with anthelmintics, particularly in school-aged children, needs to expand [102]. Mass drug administration programs with albendazole or mebendazole should be increased in the high-burden regions such as Egypt, although, because of post-treatment reinfection, there is no proof that the major soil-transmitted nematode infections will be eradicated in the near future. Mass drug administration has now ceased in Egypt due to an overall incidence of less than 1% [103], while, as reported above, schistosomiasis has been almost eradicated. Through mass drug administration with praziquantel, huge improvements have been made in the control of schistosomiasis in Egypt, with the near eradication of S. haematobium-produced bladder cancer, although recurrence is a distinct possibility, and S. mansoni infection remains endemic in the northern part of Egypt [83].

Two zoonotic helminth infections, echinococcosis and fascioliasis, are still predominant, and there are chances to exert enhanced control for them by animal treatment and, more downstream, animal vaccination to avoid transmission to humans. Special highlighting for aimed interventions is required. A global or regional strategy emphasizing Egypt for control and eradication, particularly for soil-transmitted nematode infections, filarial infections, schistosomiasis and fascioliasis, would support overall disease control in the region [64,103].

Conclusion

Zoonotic parasitic diseases remain an important health issue in Egypt. Some protozoa and helminthes are prevalent all over the country, while others have more specific geographical distribution. Animal reservoirs are varied, including domestic animals, pets, and stray and wild animals. Control measures have managed to help curb many of these zoonoses; however, additional strategies are needed to decrease their transmission country wide. Continual focused cooperation between health, veterinary, and agricultural authorities are needed to achieve high prevention and control of zoonotic parasitic infections in Egypt.

Financial support and sponsorship Nil.

Conflicts of interest

There are no conflicts of interest.

References

- Shanko K, Kemal J, Kenea D. A review on confronting zoonoses: the role of veterinarian and physician. J Veterinar Sci Technol 2015; 6:221.
- 2 Cutler SJ, Fooks AR, Van Der Poel WHM. Public health threat of new, reemerging, and neglected zoonoses in the industrialized world. Emerg Infect Dis 2010; 16:1–7.
- 3 Youssef AI, Uga S. A review of Parasitic Zoonoses in Egypt. Trop Med Health 2014; 42:3–14.
- 4 Abdel-Hafeez EH, Ahmad AK, Ali BA, Moslam FA. Opportunistic parasites among immunosuppressed children in Minia District, Egypt. Korean J Parasitol 2012; 50:57–62.
- 5 El Sahn FF, Deghedi BM, Mahdy NH, El Sahn A. The impact of intestinal parasitic infections on the nutritional status of primary school children in Alexandria, Egypt. J Egypt Public Health Assoc 1997; 72:113–151.
- 6 El-Naggar SM, el-Bahy MM, Abd Elaziz J, el-Dardiry MA. Detection of protozoal parasites in the stools of diarrhoeic patients using different techniques. J Egypt Soc Parasitol 2006; 36:487–516.
- 7 Awadallah MA, Salem LM. Zoontic enteric parasites transmitted from dogs in Egypt with special concern to *Toxocara canis* infection. Vet World 2015; 8:946–957.
- 8 El-Hakim MA, El-Sahn A. Association of parasites and diarrhoea among children less than five years of age in a rural area in Egypt. J Egypt Public Health Assoc 1996; 71:439–463.
- 9 Bakr IM, Arafa NA, Ahmed MA, Mostafa Mel H, Mohamed MK. Prevalence of intestinal parasitosis in a rural population in Egypt, and its relation to sociodemographic characteristics. J Egypt Soc Parasitol 2009; 39:371–381.
- 10 Abdel Wahed MM, Salem GH, el-Assaly TM. The role of wild rats as a reservoir of some internal parasites in Qalyobia Governorate. J Egypt Soc Parasitol 1999; 29:495–503.
- 11 El Shazly AM, Awad SE, Sultan DM, Sadek GS, Khalil HH, Morsy TA. Intestinal parasites in Dakahlia Governorate, with different techniques in diagnosing protozoa. J Egypt Soc Parasitol 2006; 36:1023–1034.
- 12 El-Sherbini GT, Gneidy MR. Cockroaches and flies in mechanical transmission of medical important parasites in Khaldyia Village, El-Fayoum, Governorate, Egypt. J Egypt Soc Parasitol 2012; 42:165–174.
- 13 Sultan K, Elmonirb W, Hegazy Y. Gastrointestinal parasites of sheep in Kafrelsheikh governorate, Egypt: Prevalence, control and public health implications. BJBAS 2016; 5:79–84.
- 14 Reeder MM, Palmer PES. Parasitic disease. In: Freeny P, Stefenson GW, editors. Margulis and Burhenne' salimentary tract radiology. 5th ed. St. Louis: Mosby 1994. 913–951
- 15 Abd El Bagi ME, Sammak BM, Mohamed AE, Al Karawi MA, Al Shahed M, Al Thagafi MA. Gastrointestinal parasite infestation. Eur Radiol 2004; 14: E116–E131.
- 16 Cuamba I, Grau-Pujol B, Nhabomba A, Gutiérrez J, Lázaro C, Mejia R. Prevalence of gastrointestinal parasites in southern Mozambique using a novel multiparallel quantitative real time PCR. BMJ Global Health 2017; 2 (Suppl 2):A61.
- 17 Zaki AM, DuPont HL, el Alamy MA, Arafat RR, Amin K, Awad MM, et al. The detection of enteropathogens in acute diarrhea in a family cohort population in rural Egypt. Am J Trop Med Hyg 1986; 35:1013–1022.
- 18 Mousa KM, Abdel-Tawab AH, Khalil HH, El-Hussieny NA. Diarrhea due to parasites particularly *Cryptosporidium parvum* in great Cairo, Egypt. J Egypt Soc Parasitol 2010; 40:439–450.
- 19 Nazeer JT, El Sayed Khalifa K, von Thien H, El-Sibaei MM, Abdel-Hamid MY, Tawfik RA, Tannich E. Use of multiplex real-time PCR for detection of common diarrhea causing protozoan parasites in Egypt. Parasitol Res 2013; 112:595–601.
- 20 El-Kadi MA, Dorrah AO, Shoukry NM. Patients with gastrointestinal complains due to enteric parasites, with reference to *Entamoeba histolytica/dispar* as detected by ELISA *E. histolytica* adhesion in stool. J Egypt Soc Parasitol 2006; 36:53–64.
- 21 Baiomy AM, Mohamed KA, Ghannam MA, Shahat SA, Al-Saadawy AS. Opportunistic parasitic infections among immunocompromised Egyptian patients. J Egypt Soc Parasitol 2010; 40:797–808.
- 22 Omran EKH, Mohammed AN. Intestinal parasites in patients with chronic abdominal pain. J Egypt Soc Parasitol 2015; 45:389–396.
- 23 Helmy MM, Abdel-Fattah HS, Rashed L. Real-time PCR/ RFLP assay to detect *Giardia intestinalis* genotypes in human isolates with diarrhea in Egypt. J Parasitol 2009; 95:1000–1004.
- 24 Foronda P, Bargues MD, Abreu-Acosta N, Periago MV, Valero MA, Valladares B, Mas-Coma S. Identification of genotypes of *Giardia*

intestinalis of human isolates in Egypt. Parasitol Res 2008; 103:1177-1181.

- 25 Nasr DS, Yousof H, Tyler K, El-Badry A, Rubio J, El-Dib N. Giardia intestinalis assemblages among Egyptian symptomatic/asymptomatic cases in Cairo. J Egypt Soc Parasitol 2018; 48:465–474.
- 26 Soliman RH, Fuentes I, Rubio JM. Identification of a novel Assemblage B subgenotype and a zoonotic Assemblage C in human isolates of *Giardia intestinalis* in Egypt. Parasitol Int 2011; 60:507–511.
- 27 Khalafalla RE. A survey study on gastrointestinal parasites of stray cats in northern region of Nile delta, Egypt. PLoS One 2011; 6:e20283.
- 28 Ghoneim NH, Abdel-Moein KA, Saeed H. Fish as a possible reservoir for zoonotic Giardia duodenalis assemblages. Parasitol Res 2012; 110:2193–2196.
- 29 Helmy YA, Krucken J, Nockler K, von Samson- Himmelstjerna G, Zessin KH. Molecular epidemiology of *Cryptosporidium* in livestock animals and humans in the Ismailia province of Egypt. Vet Parasitol 2013; 193:15–24.
- 30 Amer S, Honma H, Ikarashi M, Tada C, Fukuda Y, Suyama Y, Nakai Y. *Cryptosporidium* genotypes and subtypes in dairy calves in Egypt. Vet Parasitol 2010; 169:382–386.
- 31 Fayer R. Sarcocystis spp. in human infections. Clin Microbiol Rev 2004; 17:894–902.
- 32 El-Dakhly KM, El-Nesr KA, El-Nahass el S, Hirata A, Sakai H, Yanai T. Prevalence and distribution patterns of *Sarcocystis* spp. in buffaloes in Beni-Suef, Egypt. Trop Anim Health Prod 2011; 43:1549–1554.
- 33 El Shazly AM, Awad SE, Abdel Tawab AH, Haridy FM, Morsy TA. Echinococcosis (zoonotic hydatidosis) in street dogs in urban and rural areas, Dakahlia Governorate, Egypt. J Egypt Soc Parasitol 2007; 37:287–298.
- 34 Hamadto HA, Al FA, Farrag AB, Abdel Maksoud MK, Morsy TA. Zoonotic cutaneous leishmaniasis: reservoir host and insect vector in north Sinai, Egypt. J Egypt Soc Parasitol 2007; 37:843–850.
- 35 Shehata MG, Samy AM, Doha SA, Fahmy AR, Kaldas RM, Furman BD, Villinski JT. First report of Leishmania tropica from a classical focus of L. major in North-Sinai, Egypt. Am J Trop Med Hyg 2009; 81:213–218.
- 36 Morsy TA, Hamadto HA, Rashed SM, el-Fakahany AF, Abdalla KF. Animals as reservoir hosts for *Leishmania* in Qualyobia Governorate, Egypt. J Egypt Soc Parasitol 1990; 20:779–788.
- 37 Shehata M, el Sawaf B, el Said S, Doha S, el Hosary S, Kamal H, et al. Leishmania infantum MON-98 isolated from dogs in El Agamy, Egypt. Trans R Soc Trop Med Hyg 1990; 84:227–228.
- 38 Rosypal AC, Bowman SS, Epps SA, El Behairy AM, Hilali M, Dubey JP. Serological survey of dogs from Egypt for antibodies to *Leishmania* species.J Parasitol 2013; 99:170–171.
- 39 Montoya JG, Liesenfeld O. Toxoplasmosis. Lancet 2004; 363:1965–1976.
- 40 Elsheikha HM, Azab MS, Abousamra NK, Rahbar MH, Elghannam DM, Raafat D. Seroprevalence of and risk factors for *Toxoplasma gondii* antibodies among asymptomatic blood donors in Egypt. Parasitol Res 2009; 104:1471–1476.
- 41 Ibrahim HM, Huang P, Salem TA, Talaat RM, Nasr MI, Xuan X, Nishikawa Y. Short report: prevalence of *Neospora caninum* and *Toxoplasma gondii* antibodies in northern Egypt. Am J Trop Med Hyg 2009; 80:263–267.
- 42 Mabrouk MA, Dahawi HS. *Toxoplasma* antibodies in patients with meningoencephalitis. J Egypt Soc Parasitol 1991; 21:547–551.
- 43 Al-Kappany YM, Lappin MR, Kwok OC, Abu-Elwafa SA, Hilali M, Dubey JP. Seroprevalence of *Toxoplasma gondii* and concurrent *Bartonella* spp., feline immunodeficiency virus, feline leukemia virus, and *Dirofilaria immitis* infections in Egyptian cats. J Parasitol 2011; 97:256–258.
- 44 El-Massry A, Mahdy OA, El-Ghaysh A, Dubey JP. Prevalence of *Toxoplasma gondii* antibodies in sera of turkeys, chickens, and ducks from Egypt. J Parasitol 2000; 86:627–628.
- 45 EL-Kady GA, Makled KM, Morsy TA, Morsy ZS. Rodents, their seasonal activity, ecto- and blood-parasites in Saint Catherine area, South Sinai Governorate, Egypt. J Egypt Soc Parasitol 1998; 28:815–826.
- 46 Mazyad SA, Shoukry NM, El-Alfy NM. Efficacy of *lxodes ricinus* as a vector of zoonotic babesiosis in Sinai Peninsula, Egypt. J Egypt Soc Parasitol 2010; 40:499–514.
- 47 Haridy FM, El-Metwally MT, Khalil HH, Morsy TA. *Trypanosoma* evansi in dromedary camel: with a case report of zoonosis in greater Cairo, Egypt. J Egypt Soc Parasitol 2011; 41:65–76.
- 48 El Shazly AM, Abdel-Magied AA, El-Nahas HA, El- Metwaly MS, Morsy TA, El Sharkawy EM, Morsy AT. On the main reservoir host of *Fasciola* in Dakahlia Governorate, Egypt. J Egypt Soc Parasitol 2005; 35:243–252.

- 49 Soliman MF. Epidemiological review of human and animal fascioliasis in Egypt. J Infect Dev Ctries 2008; 2:182–189.
- 50 Dar Y, Amer S, Mercier A, Courtioux B, Dreyfuss G. Molecular identification of *Fasciola* spp. (Digenea: Fasciolidae) in Egypt. Parasite 2012; 19:177–182.
- 51 Esteban JG, Gonzalez C, Curtale F, Munoz-Antoli C, Valero MA, Bargues MD, et al. Hyperendemic fascioliasis associated with schistosomiasis in villages in the Nile Delta of Egypt. Am J Trop Med Hyg 2003; 69:429–437.
- 52 Arafa WM, Hassan AI, Snousi SA, El-Dakhly KM, Holman PJ, Craig TM, Aboelhadid SM. *Fasciola hepatica* infections in cattle and the freshwater snail *Galba truncatula* from Dakhla Oasis, Egypt. J Helminthol 2018; 92: 56–63.
- 53 Periago MV, Valero MA, El Sayed M, Ashrafi K, El Wakeel A, Mohamed MY, et al. First phenotypic description of *Fasciola hepatica/Fasciola gigantica* intermediate forms from the human endemic area of the Nile Delta, Egypt. Infect Genet Evol 2008; 8:51–58.
- 54 Mekky MA, Tolba M, Abdel-Malek MO, Abbas WA, Zidan M. Human Fascioliasis: a re-emerging disease in Upper Egypt. American J Trop Med Hyg 2015; 93:76–79.
- 55 Amer S, ElKhatam A, Zidan S, Feng Y, Xiao L. Identity of *Fasciola* spp. in sheep in Egypt. Parasit Vectors 2016; 9:623.
- 56 Mazyad SA, el-Nemr HI. The endoparasites of sheep and goats, and shepherd in North Sinai Governorate, Egypt. J Egypt Soc Parasitol 2002; 32:119–126.
- 57 Elsheikha HM, Elshazly AM. Preliminary observations on infection of brackish and fresh water fish by heterophyid encysted metacercariae in Egypt. Parasitol Res 2008; 103:971–977.
- 58 Lobna SM, Metawea YF, Elsheikha HM. Prevalence of heterophyiosis in Tilapia fish and humans in Northern Egypt. Parasitol Res 2010; 107:1029–1034.
- 59 Ibrahim MM, Soliman MF. Prevalence and site preferences of heterophyid metacercariae in Tilapia zilli from Ismalia fresh water canal, Egypt. Parasite 2010; 17:233–239.
- 60 El Khoby T, Galal N, Fenwick A. The USAID/Government of Egypt's Schistosomiasis Research Project (SRP). Parasitol Today 1998; 14:92–96.
- 61 World Health Organization. Weekly epidemiological record. 2013; 88:81-88.
- 62 El-Khoby T, Galal N, Fenwick A, Barakat R, El-Hawey A, Nooman Z, et al. The epidemiology of schistosomiasis in Egypt: summary findings in nine governorates. Am J Trop Med Hyg 2000; 62:88–99.
- 63 Hotez PJ. A handful of 'antipoverty' vaccines exist for neglected diseases, but the world's poorest billion need more. Health Aff (Millwood) 2011; 30:1080–1087.
- 64 Hotez PJ, Savioli L, Fenwick A. Neglected tropical diseases of the Middle East and North Africa: review of their prevalence, distribution, and opportunities for control. PLoS Negl Trop Dis 2012; 6:e1475.
- 65 Haggag AA, Rabiee A, Abd Elaziz KM, Gabrielli AF, Abdel Hay R, Ramzy RM. Mapping of *Schistosoma mansoni* in the Nile Delta, Egypt: assessment of the prevalence by the circulating cathodic antigen urine assay. Acta Trop 2017; 167:9–17.
- 66 Elsharazly BM, Abou Rayia DM, Antonios SN, Eissa SHH. Current status of Schistosoma mansoni infection and its snail host in three rural areas in Gharbia governorate, Egypt. Tanta Med J 2016; 44:141–150.
- **67** Mansour NS. *Schistosoma mansoni* and *Sch. Haematobium* found as a natural double infection in the Nile rat, *Arvicanthis n. niloticus*, from a human endemic area in Egypt. J Parasitol 1973; 59:424.
- 68 Haridy FM, Ibrahim BB, Morsy TA, Ramadan NI. Human taenaisis and cysticercosis in slaughtered cattle, buffaloes and pigs in Egypt. J Egypt Soc Parasitol 1999; 29:375–394.
- 69 Abdel-Hafeez EH, Kamal AM, Abdelgelil NH, Abdel-Fatah M. Parasites transmitted to human by ingestion of different types of meat, El-Minia City, El-Minia Governorate, Egypt. J Egypt Soc Parasitol 2015; 45:671–680.
- 70 Khalil HM, el Shimi S, Sarwat MA, Fawzy AF, el Sorougy AO. Recent study of *Hymenolepis nana* infection in Egyptian children. J Egypt Soc Parasitol 1991; 21:293–300.
- 71 Safar HH, Eldash HH. Parasitic infections: is male and female difference for anemia and growth retardation evident? J Egypt Soc Parasitol 2015; 45:467–475.
- 72 Sadjjadi SM. Present situation of echinococcosis in the Middle East and Arabic North Africa. Parasitol Int 2006; 55:S197–S202.
- 73 Kandeel A, Ahmed ES, Helmy H, El Setouhy M, Craig PS, Ramzy RM. A retrospective hospital study of human cystic echinococcosis in Egypt. East Mediterr Health J 2004; 10:349–357.

- 74 Dyab KA, Hassanein R, Hussein AA, Metwally SE, Gaad HM. Hydatidosis among man and animals in Assiut and Aswan Governorates. J Egypt Soc Parasitol 2005; 35:157–166.
- 75 Aaty HE, Abdel-Hameed DM, Alam-Eldin YH, El-Shennawy SF, Aminou HA, Makled SS, Darweesh SK. Molecular genotyping of *Echinococcus granulosus* in animal and human isolates from Egypt. Acta Trop 2012; 121:125–128.
- 76 Abdel-Moein KA, Hamza DA. Norway rat (*Rattus norvegicus*) as a potential reservoir for *Echinococcus granulosus*: a public health implication. Acta Parasitol 2016; 61:815–819.
- 77 EI-Dib NA, EI-Badry AA, Ta-Tang TH, Rubio JM. Molecular detection of *Capillaria philippinensis*: an emerging zoonosis in Egypt. Exp Parasitol 2015; 154:127–133.
- 78 Attia RA, Tolba ME, Yones DA, Bakir HY, Eldeek HE, Kamel S. Capillaria philippinensis in Upper Egypt: has it become endemic? Am J Trop Med Hyg 2012; 86:126–133.
- **79** Okulewicz Z, Zalesny G. Biodiversity of Capillariinae. Wiad Parazytol 2005; 51:9–14.
- 80 Shalaby HA, Abdel-Shafy S, Derbala AA. The role of dogs in transmission of Ascaris lumbricoides for humans. Parasitol Res 2010; 106:1021–1026.
- 81 Abdou Nel S, Dronen NO. Studies on the juveniles of a species of *Anisakis* (Nematoda: Anisakidae) from the orangespotted trevally, Carangoides bayad (Carangidae), from the Red Sea, Egypt. J Egypt Soc Parasitol 2007; 37:1055–1064.
- 82 Bahgat MA, El Gindy AE, Mahmoud LA, Hegab MH, Shahin AM. Evaluation of the role of *Ancylostoma caninum* in humans as a cause of acute and recurrent abdominal pain. J Egypt Soc Parasitol 1999; 29:873–882.
- 83 Salem S, Mitchell RE, El-Alim A, El-Dorey A, Smith JA, Barocas DA. Successful control of schistosomiasis and the changing epidemiology of bladder cancer in Egypt. BJU Int 2011; 107:206–211.
- 84 Antonios SN, Eid MM, Khalifa EA, Othman AA. Sero prevalence study of *Toxocara canis* in selected Egyptian patients. J Egypt Soc Parasitol 2008; 38:313–318.
- 85 El Shazly AM, Mohammed RM, El-Beshbishi SN, Azab MS, El-Ghareeb AS, Abdeltawab AH, Zalook TK. Soil parasites particularly *Toxocara* eggs in Egypt. J Egypt Soc Parasitol 2009; 39:151–162.
- 86 EI-Tras WF, Holt HR, Tayel AA. Risk of *Toxocara canis* eggs in stray and domestic dog hair in Egypt. Vet Parasitol 2011; 178:319–323.
- 87 Siam MA, Michael SA, Ghoneim NH. Studies on the isolation of the infective stages of *Trichinella spiralis* and *Toxoplasma gondii* from fresh and processed pork in Egypt. J Egypt Public Health Assoc 1979; 54:163–170.
- 88 Pozio E. Current status of food-borne parasitic zoonoses in Mediterranean and African regions. Southeast Asian J Trop Med Public Health 1991; 22:85–87.
- 89 Loutfy NF, Awad OM, El-Masry AG, Kandil GM. Study on rodents infestation in Alexandria and prevalence of *Trichinella spiralis* infection among them. J Egypt Soc Parasitol 1999; 29:897–909.
- 90 Weil GJ, Ramzy RM, El Setouhy M, Kandil AM, Ahmed ES, Faris R. A longitudinal study of Bancroftian filariasis in the Nile Delta of Egypt: baseline data and one-year follow-up. Am J Trop Med Hyg 1999; 61:53–58.
- 91 Hassan M, Sanad MM, el-Karamany I, Abdel-Tawab M, Shalaby M, el-Dairouty A, et al. Detection of DNA of W. bancrofti in blood samples by QC-PCR-ELISA-based. J Egypt Soc Parasitol 2005; 35:963–970.
- 92 Kamal IH, Fischer P, Adly M, El Sayed AS, Morsy ZS, Ramzy RM. Evaluation of a PCR-ELISA to detect *Wuchereria bancrofti* in *Culex pipiens* from an Egyptian village with a low prevalence of filariasis. Ann Trop Med Parasitol 2001; 95:833–841.
- 93 Makhlouf LM, Monib ME, Abou-Zkam AA, Romia SA, el-Ganayni GA, Handousa A. A microfilaria in a stray cat from Assiut, Egypt. J Egypt Soc Parasitol 1989; 19:247–249.
- 94 Abdel-Rahman SM, Mahmoud AE, Galal LA, Gustinelli A, Pampiglione S. Three new cases of human infection with *Dirofilaria repens*, one pulmonary and two subcutaneous, in the Egyptian governorate of Assiut. Ann Trop Med Parasitol 2008; 102:499–507.
- 95 Maher M, El-Dib N, Zakaria E. Dirofilaria conjunctivae in eyelid nodule. In: Kamel R, Lumley J. Textbook of tropical surgery. UK: Westminister Publishing Limited. 2004. 1267–1268.
- 96 Fuller LC. Epidemiology of scabies. Curr Opin Infect Dis 2013; 26:123–126.
- 97 Anbar T, El-Domyati M, Mansour H, Ahmad H. Scaly scalp associated with crusted scabies: case series. Dermatol Online J 2007; 13:18.
- 98 Felt SA. Acariasis in Captive Fat-Tailed Jirds (*Pachyuromys duprasi*). J Zoo Wildl Med 2009; 40:217–219.

- **99** Yassin M. Mange mites causing scabies in Egyptian buffaloes at Giza Governorate. Egypt J Egypt Soc Parasitol 2011; 41: 55-64.
- 100 Mazyad SA, Abel El-Kadi M. Ornithonyssus (Acari: Macronyssidae) mite dermatitis in poultry field-workers in Almarg, Qalyobiya governorate. J Egypt Soc Parasitol 2005; 35:213–222.
- 101 Shoukry NM, El-Naggar MH, Darwish AB, Soliman BA, El-Sawaf BM. Studies on rodents role as reservoir hosts of leishmaniasis with specical

reference to their ectoparasites in Suez Governorate. J Egypt Soc Parasitol 2006; 36:93–106.

- 102 World Health Organization. Soil-transmitted helminthiasis. Weekly Epidemiol Rec 2008; 83:237–252.
- 103 World Health Organization. Global Programme to Eliminate Lymphatic Filariasis. Progress Report 2000–2009 and Strategic Plan 2010–2020; Preventive Chemotherapy and Transmission Control (PCT), Department of Control of Neglected Tropical Diseases 2010.